Plasma Leptin Is Related to Proinflammatory Status and Dietary Intake in Patients with Chronic Obstructive Pulmonary Disease

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Recent studies suggest that leptin, a protein synthesized by adipose tissue and encoded by the ob gene (1), plays an important role in the energy balance. Leptin is postulated to represent the afferent hormonal signal to the brain including the hypothalamus in a feedback mechanism regulating the fat mass (FM). Leptin binds to the leptin receptor (2) in the hypothalamus (3), the brain nucleus that plays a central role in the regulation of feeding behavior and energy balance. In animal models, the result of this interaction is a decrease in food intake (4). There is now evidence that the effects of leptin on food intake are mediated by two limbs of the weight control system: the appetite-stimulating peptide, neuropeptide Y, and the satiety-stimulating, melanocyte-stimulating hormone (5). Furthermore, leptin may mediate energy expenditure by both the satiety-stimulating, melanocyte-stimulating hormone (5). Furthermore, leptin may mediate energy expenditure by both the satiety-stimulating, melanocyte-stimulating hormone (5).

Leptin is exponentially related to FM in emphysema (r = 0.74, p < 0.001) and in chronic bronchitis (r = 0.80, p = 0.001). Furthermore, a significant partial correlation coefficient between leptin and sTNF-R55 adjusted for FM and oral corticosteroid use was seen in emphysema (r = 0.81, p < 0.001) but not in chronic bronchitis. In 17 predominantly emphysematous depleted male patients with COPD, baseline plasma leptin divided by FM was in addition logarithmically inversely related to baseline dietary intake (r = −0.50, p = 0.047) and to the degree of weight change after 8 wk of nutritional support (r = −0.60, p = 0.017). This proposed cytokine-leptin link in pulmonary cachexia may explain the poor response to nutritional support in some of the cachectic patients with COPD and may open a novel approach in combating this significant comorbidity in COPD. Schols AMWJ, Creutzberg EC, Buurman WA, Campfield LA, Saris WHM, Wouters EFM. Plasma leptin is related to proinflammatory status and dietary intake in patients with chronic obstructive pulmonary disease.
(TNF-α) in patients with COPD suffering from weight loss (13, 14). Clinical data regarding the possible involvement of leptin in the pathophysiology of inflammation-associated chronic wasting are however lacking.

The present study was undertaken to investigate cross-sectionally the relationship between plasma leptin concentrations and soluble TNF receptor (sTNF-R) 55 and 75 concentrations, as a reflection of an enhanced inflammatory status, in patients with COPD stratified into emphysema and chronic bronchitis. Furthermore, in a group of depleted patients with COPD it was prospectively studied if leptin was a determining factor of dietary intake, resting energy expenditure (REE), and the response to nutritional therapy.

METHODS

Patients

Cross-sectional study. A random group of patients with COPD, consecutively admitted to a pulmonary rehabilitation center, were included in the study when they fulfilled the following criteria: (1) COPD according to the American Thoracic Society guidelines (15) and chronic airflow obstruction defined as a measured forced expiratory volume in one second (FEV₁) less than 70% of the reference value; (2) irreversible obstructive airway disease, i.e., <10% improvement in FEV₁, expressed as percentage of predicted after inhalation of a β₂-agonist; (3) in clinically stable condition, not suffering from a respiratory tract infection; (4) no concomitant confounding diseases, such as malignant disease, gastrointestinal disorders, severe endocrine disorders or recent surgery; (5) no suspected abnormal fluid balance as manifested by the presence of edema or regular use of diuretics. In order to increase homogeneity of the study population, only male subjects were included.

Prospective study. The second group consisted of male patients with COPD fulfilling the same inclusion criteria as indicated in the cross-sectional study, and suffering from below normal body weight (body mass index [BMI]; body weight/height²) ≤ 23 kg/m² and/or depletion of fat-free mass (fat-free mass index [FFM]; FFM/height²) ≤ 16 kg/m². These patients received standardized nutritional therapy consisting of 500 to 750 kcal/d given as three liquid supplements of 200 ml each, as an integrated part of an in-patient pulmonary rehabilitation program. Response to nutritional therapy was defined as the weight change reached after 8 wk of treatment.

The study was approved by the medical ethical committee of the University Hospital of Maastricht. Informed consent was obtained from all subjects. All measurements were performed during the first 2 wk after admission to the center, and in the prospective study in addition after 8 wk of nutritional treatment.

Body Composition

Body height was determined to the nearest 0.5 cm (Lameris WM 715; Breukelen, The Netherlands) with subjects standing barefoot. Body weight was measured with a beam scale to the nearest 0.1 kg (SECA, Breukelen, The Netherlands) with subjects standing barefoot. Body height was determined to the nearest 0.5 cm (Lameris WM 715; Lameris WM 715; Lameris WM 715; Lameris WM 715; Lameris WM 715). Body height was determined to the nearest 0.5 cm (Lameris WM 715; Lameris WM 715; Lameris WM 715; Lameris WM 715; Lameris WM 715). Body height was determined to the nearest 0.5 cm (Lameris WM 715; Lameris WM 715; Lameris WM 715; Lameris WM 715; Lameris WM 715).

To measure total body water (TBW) each patient received a weighed (1 g/L estimated TBW) oral dose of deuterium-labeled water (D₂O; 99.84 atom percentage excess) mixed into 70 ml water in the late evening around 10:00 p.m. Just before and approximately 10 h later, after complete emptying of the bladder, a urine sample was obtained. Urine was analyzed for deuterium with an isotope ratio mass spectrometer (16). Deuterium dilution space was calculated from the quantity of administered D₂O and the urine D₂O concentrations after complete distribution. TBW was calculated from these values by applying a conversion factor of 1.04. This correction accounts for the exchange of labile hydrogen which occurs in humans during the equilibration period. FFM was calculated assuming a hydration factor of 0.73. FM was calculated by subtracting FFM from body weight.

In order to assess possible disturbances in body fat distribution in the cross-sectional study group, subregional FM was assessed by dual-energy X-ray absorptiometry (DXA), which is a direct method of assessing bone mineral content and the soft tissue surrounding the bone (FFM and FM). Each patient, lying in supine position on a scan table for approximately 15 min, was scanned by a DPX-L Bone densitometer (Lunar Radiation Corp., Madison, WI; voltage: 76.0 kVp, current 150 μA, collimation 1.68 mm). Multiple fast-speed transverse scans from head to toes with 1-cm intervals were performed, with a scan area of 576 × 1,986 mm and a sample interval of 1/32. A rectilinear scanner was used to detect density differences as the two concentrations of photon energy were projected through the subject. The scanner used a constant potential X-ray source at 78 kV and a K-edge filter to achieve a congruent beam of stable dual-energy radiation with effective energies of 38 and 70 keV. Data were collected in maximal 205 scan lines × 120 sample points (pixel size 4.8 × 9.6 mm). E tranece radiation dose was minimal (<0.02 mSv/scan) (17). Total and subregional FM were derived according to computer algorithms (Lunar software version 1.3; Madison, WI) provided by the manufacturer.

Resting Energy Expenditure

REE was measured by an open-circuit indirect calorimetry system using a ventilated hood (Oxycon; Jaeger, Wurzburg, The Netherlands) (18). Measurements were started in the early morning (8:30 ± 0.5 h). Patients were in a fasting state for at least 10 h and had a period of at least 30 min bed rest prior to the measurement during which subjects were comfortably lying on a bed in supine position. After stabilization, REE was recorded during a period of 20 min and calculated from oxygen consumption and carbon dioxide production using the abbreviated Weir formula.

Dietary Intake

In the cross-sectional study, dietary intake was assessed using the dietary history method with cross check. In the prospective study, food intake was recorded during 4 d before the start of the nutritional intervention and the mean intake of the 4 d was taken for analysis. The information was coded for computer nutrient analysis by the same trained dietician. The nutrient data base was derived from the Dutch food composition tables (19).

Lung Function

Lung function testing included spirometry (FEV₁ and FVC), thoracic gas volumes (TLC) and diffusing capacity for carbon monoxide (DLCO; M asterlab; Jaeger, Wurzburg, FRG). Lung function values were expressed as a percentage of predicted (20). Blood was drawn from the brachial artery at rest while breathing room air. A retial pressures of oxygen and carbon dioxide (Pao₂, Pco₂) were analyzed on a blood gas analyzer (ABL 330; Radiometer, Copenhagen).

Assessment of Emphysema

Evaluation of the presence and severity of emphysema was performed by high-resolution computed tomography (HRCT) using a commercial scanner (Somaton Plus; Siemens, Erlangen, FRG; voltage: 137 kVp; current: 220 mA; collimation: 1.0 mm; scanning time: 10 s). Five thin-section CT scans were obtained with the patient supine during breath hold at end-expiration; two scans of the upper and two scans of the lower lung zones at 3 and 6 cm above and below the carina and one scan at the carina. Images were made at a concentration of 800 Hounsfield units (HU) and window width of 1,600 HU, which is appropriate for lung detail. The severity and extent of emphysema of each scan was visually scored on a four-point scale independently by two observers according to the direct observational method of Sakai (21). For each of the lung sections, the score for the severity of emphysema was multiplied by the score for the extent; the resultant scores were subsequently summed to give a total HRCT score. Visual scores ranged from 0 (no emphysema) to 120 (severe emphysema). Patients with a visual score >30 were subtyped as chronic bronchitis and patients with a visual score ≤30 were subtyped as emphysema (22).

HRCT is a sensitive technique for the evaluation of the presence and severity of emphysema; in patients with emphysema the densitometric parameters substantially differ from the corresponding values in patients with chronic bronchitis and healthy control subjects, regardless of the level of inspiration (23).
Collection and Analysis of Plasma Samples
From all patients, blood was obtained in the fasting state by venipuncture at 9:00 a.m. Blood was collected in evacuated blood collection tubes (Sherwood Medical, St. Louis, Mo.) containing ethylenediaminetetraacetic acid (EDTA). Plasma was separated from blood cells by centrifugation at 1,000 g for 10 min at 4°C within 1 h after collection. Plasma samples were stored at −70°C until analysis. sTNF-R55 and -R75 were measured using specific sandwich ELISA described elsewhere (24). In short, monoclonal antibodies (mAb) M R1-1 and M R2-2 were coated on immunosassay plates (Nunc-Immuno Plate Maxisorp, Roskilde, Denmark). The standards used were recombinant human sTNF-R55 and sTNF-R75. Specific biotin-labeled polyclonal rabbit anti-human sTNF-R IgG were used as detector reagents followed by streptavidin-peroxidase conjugate (Dako, Glostrup, Denmark). Photospectrometry (450 nm) was performed using a micro-ELISA auto-reader. The detection limit of both assays was 100 pg/ml. Leptin concentrations were measured using a double antibody sandwich ELISA assay using a mAb specific for human leptin. The lower concentration of detection was 0.25 ng/ml and the upper limit 50 ng/ml. The intra-assay using a mAb specific for human leptin. The lower concentration of detection was 0.25 ng/ml and the upper limit 50 ng/ml. The intra- and interassay variation were 9% and 12% respectively. The leptin concentrations of normal-weight healthy subjects ranged from 1 to 12 ng/ml.

Fasting serum concentration of glucose was determined by spectrophotometric analysis (Cobas Mira; Hoffmann-La Roche, Basel, Switzerland).

Statistical Analysis
Results are given as mean ± SD. Differences between groups were statistically analyzed using an unpaired Student’s t test. In the patients with leptin values below the detection limit (0.25 ng/ml), the value 0.25 ng/ml is used in the analysis. A t test curve estimation, linear, exponential or logarithmic Pearson product moment correlation coefficients were calculated. The relationship between leptin and the sTNF receptors was adjusted for FM and oral corticosteroid use using partial correlation analysis. After the simple correlations, a regression model was fitted to the data to select the variables that contributed to the explained variation in plasma leptin concentration. Significance was determined at the 5% level. Data were analyzed according to the guidelines of Altman and coworkers (25), using SPSS (Statistical Package for the Social Sciences, version 6.0 for Windows, SPSS Inc., Chicago, IL).

RESULTS
Cross-sectional Study
Characteristics of the study group stratified into the COPD subtypes (27 patients with emphysema, 15 patients with chronic bronchitis) are given in Table 1. The total group was characterized by severe lung function impairment. Patients with emphysema were characterized by a significantly lower FEV₁, DLCO, and P A O₂ compared with those with chronic bronchitis. Emphysematous patients expressed also a significantly lower BMI owing to a significantly lower FM (mean difference 7.8 kg; 95% confidence interval [CI]: 3.2 to 12.4 kg), whereas the groups were not different in FFM and in the prevalence of recent weight loss (emphysema: 14/27 versus chronic bronchitis: 6/15, p = 0.340). Dietary intake was nearly significantly higher in the patients with the emphysematous subtype of COPD compared with the patients with the bronchitic subtype (p = 0.055), whereas no difference was seen in REE.

Maintenance medication in the majority of patients consisted of theophylline (inhaled) B₂-agonists, and inhaled corticosteroids. Furthermore, 11 of 27 patients with emphysema versus eight of 15 patients with chronic bronchitis were on low-dose systemic corticosteroids (prednisone ≤ 10 mg/d; p = 0.322). No differences were seen in serum glucose between the patients with emphysema and chronic bronchitis (Table 1). On DXA -analysis, visceral FM estimated by trunk FM expressed as percentage of total FM was higher in patients with emphysema compared with chronic bronchitis (52.1 ± 5.4% versus 47.2 ± 6.7%, p = 0.023). However, no influence of oral corticosteroid use on the distribution of body fat could be established.

As expected based on the lower FM, mean detectable plasma leptin was significantly lower in the patients with emphysema compared with patients with chronic bronchitis (Table 1). Nondetectable concentrations (below 0.25 ng/ml) were found in eight of 27 patients with emphysema relative to two of 15 patients with chronic bronchitis. A large interindividual variation was seen, which is illustrated by the distribution of FM among the patients with leptin concentrations below the detection limit (Figure 1). The curvilinear correlation coefficient between leptin and FM was 0.74 (p < 0.001) in the patients with emphysema (Figure 1A) and 0.80 (p = 0.001) in the patients with chronic bronchitis (Figure 1B). In addition, significant correlation coefficients between leptin and visceral FM expressed as percentage of total FM were revealed in the emphysematous patients (r = 0.45, p = 0.030) and in the bronchitic patients (r = 0.77, p = 0.003). The relation between leptin and FM or proportion of visceral FM was not affected by oral corticosteroid use.

After adjustment for FM and oral corticosteroid use as possible confounders, a significant partial correlation coefficient was found in the total group between leptin and sTNF-R55 (r = 0.59, p < 0.001) but not between leptin and sTNF-R75 (r = −0.08, p = 0.633). Figure 2 shows a striking difference in the relationship between sTNF-R55 and leptin concentration between the COPD subtypes. Whereas in the patients with emphysema a highly significant partial correlation coefficient was found between sTNF-R55 and leptin after adjustment for FM and oral corticosteroid use (r = 0.81, p < 0.001; Figure 2A), such a relationship was not found in chronic bronchitis (r = 0.29, p = 0.369; Figure 2B).

There are minor discrepancies in the number of subjects in Figures 1A and 2A and 1B and 2B. These discrepancies can be explained by the fact that in two of 27 patients with emphysema and in one of 15 patients with chronic bronchitis FM was not measured.

On stepwise regression analysis, FM and sTNF-R55 significantly explained 64% of the variation in plasma leptin concentration (p < 0.001). HRCT score as a measure of the extent of

<table>
<thead>
<tr>
<th>TABLE 1</th>
<th>PATIENT CHARACTERISTICS</th>
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<tbody>
<tr>
<td></td>
<td>Emphysema (n = 27)</td>
</tr>
<tr>
<td></td>
<td>(mean ± SD)</td>
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<tr>
<td>Age, yr</td>
<td>67 ± 8</td>
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<tr>
<td>BMI, kg/m²</td>
<td>21.6 ± 3.0</td>
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<tr>
<td>FM, kg</td>
<td>47.0 ± 6.7</td>
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<tr>
<td>FM, kg</td>
<td>15.7 ± 6.4</td>
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<tr>
<td>Dietary intake, kcal/24 h</td>
<td>2,144 ± 536</td>
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<tr>
<td>REE, kcal/24 h</td>
<td>1,494 ± 194</td>
</tr>
<tr>
<td>Pvc, % pred</td>
<td>86 ± 14</td>
</tr>
<tr>
<td>FEV₁, % pred</td>
<td>33 ± 10</td>
</tr>
<tr>
<td>TLC, % pred</td>
<td>120 ± 19</td>
</tr>
<tr>
<td>D CO₂, % pred</td>
<td>45 ± 17</td>
</tr>
<tr>
<td>P A O₂, kPa</td>
<td>9.4 ± 1.4</td>
</tr>
<tr>
<td>P A CO₂, kPa</td>
<td>5.8 ± 0.7</td>
</tr>
<tr>
<td>sTNF-R55, ng/ml</td>
<td>0.65 ± 0.32</td>
</tr>
<tr>
<td>sTNF-R75, ng/ml</td>
<td>1.57 ± 0.39</td>
</tr>
<tr>
<td>Glucose, mmol/L</td>
<td>5.9 ± 0.9</td>
</tr>
<tr>
<td>Leptin, ng/ml</td>
<td>2.6 ± 2.7</td>
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*p Mean value of detectable leptin concentrations: n = 19 (emphysema) and n = 13 (bronchitis) (lower detection limit of the assay: 0.25 ng/ml.)
emphysema, FFMI as a measure of functional tissue depletion, and oral corticosteroid use were excluded from the model (Table 2).

**Prospective Study**

Prospectively, 17 male patients with COPD suffering from depletion of body weight and/or FFM were studied. In this group, mean age (± SD) amounted to 65 ± 7 yr, BMI 19.4 ± 2.1 kg/m², FFM 44.5 ± 5.0 kg, FM 13.9 ± 6.5 kg, FEV₁ 30 ± 10% pred, and DLCO 37 ± 14% pred. In these patients the DLCO, as a less precise measure of the extent of emphysema, was comparable to the DLCO of the patients with emphysema described in the cross-sectional study and significantly lower than in the bronchitic patients (p < 0.05). It is known that HRCT assessment of emphysema highly correlates with the DLCO. In the study of Lamers (26) a high prevalence (93%) of HRCT-scored emphysema was observed in patients with a DLCO < 50% of predicted and a low prevalence (19%) in patients with a DLCO ≥ 50% of predicted. Furthermore, DLCO (percentage of predicted) significantly inversely correlated with the HRCT emphysema score (26). Mean weight gain after 8 wk of nutritional supplementation was 2.9 ± 2.7 kg, consisting of 1.6 ± 2.0 kg FFM.

In most patients (10 of 17) leptin concentrations were undetectable. Serum glucose amounted to 6.2 ± 1.0 mmol/L mean ± SD. Similar to the cross-sectional study, maintenance medication in the majority of patients consisted of theophylline (inhaled) β₂-agonists, and inhaled corticosteroids. Furthermore, nine of 17 patients were using low-dose systemic corticosteroids (prednisone ≥ 10 mg/d). No differences in leptin, sTNF receptors, or glucose were seen between patients who were receiving oral corticosteroids and patients who were not.

Figure 3 shows that mean dietary intake before the start of the nutritional intervention was logarithmically inversely related to baseline leptin concentration divided by FM (r = −0.50, p = 0.047) while no relationship was found between REE and leptin. Baseline leptin concentration, also divided by FM, was in turn logarithmically inversely related to the body weight change (but not specifically to the changes in FFM or FM) reached after 8 wk of nutritional intervention (r = −0.60, p = 0.017; Figure 4).

The discrepancies in the number of subjects in Figures 3 and 4 can be addressed to the fact that in one of 17 patients dietary intake was not assessed and in two of 17 patients the change in body weight was not measured.

**DISCUSSION**

This is the first clinical study showing that enhanced concentrations of leptin are related to proinflammatory status in patients with COPD. A significant relationship between plasma concentrations of leptin and sTNF-R55 adjusted for fat mass and oral corticosteroid use was found, particularly in the emphysematous subtype. Subsequently in a group of depleted emphysematous patients with COPD, baseline leptin concentration was in turn inversely related to baseline dietary intake.
as well as to the weight change after 8 wk of nutritional intervention.

Leptin was found to be associated with FM in line with the reported feedback mechanism involved in the regulation of FM. Because plasma leptin was unrelated to FFMI (as a measure of functional tissue depletion), it appears that this feedback mechanism does not consider global body composition in patients with COPD. We were however not able to compare leptin levels in COPD with normal leptin levels, because we did not include a healthy age-matched control group. The difference in plasma leptin between emphysema and chronic bronchitis was expected because of significant differences in FM. The lower leptin levels in emphysema could be related to the higher dietary intake revealed in this subgroup, according to the normal feedback mechanism of fat mass by leptin. The fact that dietary intake was increased in the patients with emphysema, despite a similar REE and prevalence of recent weight loss compared with the patients with chronic bronchitis, could be explained by an increased “wasting” of energy related to a previously reported increased activity-related energy expenditure (27).

The fact that FM could only partly explain the variation in leptin concentrations suggests that other factors might be involved. In this study we identified a significant relationship between leptin and sTNF-R55 concentrations in patients with COPD after adjustment for FM and oral corticosteroid use. Further indications for involvement of inflammation in the pathogenesis of weight loss in COPD are given by observations of others. Using an immunoradiometric assay, di Francia and coworkers demonstrated elevated TNF-α concentrations in serum of patients with COPD suffering from weight loss compared with weight-stable patients (13). In contrast de Godoy, using an ELISA assay which measured only biologically active TNF-α, did not find differences in TNF-α serum concentrations between weight-losing and weight-stable patients with COPD (14). The TNF-α production of peripheral monocytes of patients with recent weight loss was however enhanced after stimulation with lipopolysaccharide (LPS) when compared with weight-stable patients with COPD and control subjects (14).

The reason for the discrepancy between the correlations between leptin and sTNF-R55 and leptin and sTNF-R75 is unclear, but other studies from our group have shown similar results. In lung cancer, patients who exhibited a weight loss >10% tended to exhibit a higher level of sTNF-R55 compared with patients with weight loss <10% (p = 0.06), whereas there was no difference in sTNF-R75 levels. In accordance, the percentage of weight loss was significantly correlated with sTNF-R55 (r = 0.59, p = 0.02) but not with sTNF-R75 (28). In another study in lung cancer, plasma sTNF-R55 was significantly higher in patients with a REE ≥110% of the Harris and Benedict prediction equations and suffering from recent weight loss, whereas plasma sTNF-R75 was not different (29). So it seems that sTNF-R55 and sTNF-R75 exert some differential effects on weight maintenance and energy balance; sTNF-R55 has shown to be related to both dietary intake and metabolic parameters such as weight loss and REE, whereas sTNF-R75 has not. In addition, sTNF-R55 may be more sensitive to metabolic changes than sTNF-R75.

Experimental animal studies have provided evidence for a link between proinflammatory cytokines and leptin. Cytokine treatment (TNF-α, interleukin-1 [IL-1]) of fasted hamsters increased concentrations of leptin in the circulation and leptin messenger RNA (mRNA) in adipose tissue. The increase in circulating leptin concentrations correlated with a decrease in food intake (11). The hypothesis that cytokine induction of leptin may play a significant role in the anorexia and cachexia of inflammatory diseases was further illustrated by Sarraf showing that administration of the proinflammatory cytokines TNF-α, IL-1, and to a lesser extent leukemia inhibitory factor produced a prompt and dose-dependent increase in serum leptin and leptin mRNA expression in the adipose tissue of mice. In contrast, the cytokines IL-10, IL-4, and ciliary neurotrophic factor, not known to induce anorexia or a decrease in intake, did not affect leptin gene expression or serum leptin concentrations (12). No data are available yet on leptin mRNA expression in FM of patients suffering from cachexia.

### Table 2

<table>
<thead>
<tr>
<th>Cumulative R²</th>
<th>p Value</th>
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<tr>
<td>FM, kg</td>
<td>0.46</td>
</tr>
<tr>
<td>sTNF-R55, ng/ml</td>
<td>0.64</td>
</tr>
<tr>
<td>HRCT score, 0–120</td>
<td>0.728</td>
</tr>
<tr>
<td>FFMI, kg/m²</td>
<td>0.717</td>
</tr>
<tr>
<td>Oral corticosteroid use, no/yes</td>
<td>0.432</td>
</tr>
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</table>

Definition of abbreviation: R² = coefficient of determination.

**Figure 3.** Inverse relationship between dietary intake before nutritional therapy and baseline plasma leptin concentration divided by fat mass in depleted patients with COPD (r = −0.50, p = 0.047).

**Figure 4.** Inverse relationship between the change in body weight after 8 wk of nutritional therapy and baseline plasma leptin concentration divided by fat mass in depleted patients with COPD (r = −0.60, p = 0.017).
Furthermore, it should be pointed out that leptin gene expression may be different between rodents and humans.

On the opposite end of the energy balance spectrum, recent data have suggested a key role for TNF-α in the insulin resistance of obesity and of noninsulin-dependent diabetes mellitus (NIDDM) (30). In humans, a strong positive correlation was found between the degree of obesity based on the BMI, hyperinsulinemia, and relative TNF-α mRNA concentrations in adipose tissue (31). The results of another recent study did however not reveal a significant relationship between individual insulin sensitivity on the one hand and circulating concentrations of TNF-α and leptin on the other in patients with offspring NIDDM (32). No significant relationship was found between leptin and TNF-α in these patients in contrast to the findings of our group. Based on the present study, we cannot exclude an influence of alterations in insulin sensitivity, possibly induced by oral corticosteroid therapy, on the observed relationship between sTNF receptors and leptin in patients with COPD. Jakobson and coworkers reported nevertheless that insulin resistance was not exhibited by patients with advanced COPD compared with healthy subjects (33).

A substantial proportion of patients used oral corticosteroids as maintenance medication. Reports concerning the effects of oral corticosteroid use on leptin are contradictory. Two days of oral corticosteroid administration (dexamethasone 1.5 mg/d) in healthy subjects resulted in significantly increased serum concentrations of leptin, owing to enhanced leptin messenger RNA concentrations, and of insulin, but not of serum glucose (34). In addition, another study reported that administration of dexamethasone during 4 d (2.5 mg/d) to healthy lean and obese subjects induced a significant increase in plasma leptin concentrations. Furthermore, significant correlations between the change in plasma leptin on the one hand and BMI, baseline plasma leptin, and plasma dexamethasone concentrations on the other were revealed (35). In lean healthy male volunteers, Tataranni investigated whether acute intravenous administration of glucocorticosteroids (methylprednisolone 125 mg) or prolonged oral treatment (40 mg/d during 4 d) affected plasma leptin concentrations. A cute administration had no effects on insulin, free fatty acids (FFA), or leptin concentrations as compared with placebo, whereas prolonged administration significantly increased fasting concentrations of insulin, but not of glucose, FFA, or leptin (36). In the cross-sectional part of this study we found no influence of prolonged oral corticosteroid use on the relation of leptin with FM or with sTNF-R55. Aiso concentrations of leptin and sTNF receptors were comparable between patients using prednisone or not. In the prospective study, no differences in leptin, sTNF receptors, or glucose were seen between patients who were receiving oral corticosteroids and patients who were not.

A nother well known effect of chronic use of glucocorticoids is shifting in body fat distribution toward a higher visceral fat compartment. Visceral adiposity has furthermore been associated with increased serum leptin concentrations in healthy male subjects, in contrast to females (37). In the cross-sectional study indeed a significant, positive relationship was established between the proportion of visceral fat and leptin, but no influence of chronic oral corticosteroid use on fat distribution was seen.

Overall, on the basis of the present study, the impact of the presumed effects of oral corticosteroids on leptin metabolism and on the relationship between leptin and sTNF receptors appears marginal.

The cause of the systemic inflammation in patients with COPD is unknown. In our study group the relation between leptin and sTNF-R55 was stronger in the patients with emphysema than in those with chronic bronchitis. This difference could be related to a larger proportion of depleted patients or to other factors in the pathophysiology of the disease such as chronic or intermittent hypoxemia. Ghezzi and coworkers previously demonstrated that LPS-stimulated human monocytes increased their release of TNF-α and IL-1 during hypoxia (38). More recently, Hempel and coworkers showed that hypoxia also caused significant changes in the LPS-stimulated release of the cytokines TNF-α and IL-1β by the human alveolar macrophage. These changes could be mediated by an altered synthesis of anti-inflammatory prostaglandins (PG), e.g., PGE₂, or by a direct effect of hypoxemia on gene regulation caused by changes in cell oxidant tone (39). In addition, in patients with chronic bronchitis, the FM was significantly higher than in emphysema, so in this subgroup of patients the relation between FM, leptin, and dietary intake is probably less disturbed by the presence of a systemic inflammatory response. We recognize, however, that the relationship between leptin and systemic inflammation may also be related to the nutritional status which was better in the bronchitic patients than in the emphysematous patients.

The results of this study may have important therapeutic implications. Despite an overall positive effect of nutritional support on body composition and functional performance in depleted patients with COPD, we and others have reported that a proportion of the patients did not respond to this treatment (40). Based on the present findings, it may be argued whether oral nutritional support alone is an appropriate treatment strategy in depleted patients with COPD suffering from anorexia owing to the presence of a systemic inflammatory response. In line with this hypothesis we showed in a subgroup of predominantly emphysematous depleted patients with COPD that baseline plasma leptin concentrations were inversely related to baseline dietary intake and to the change in body weight after 8 wk of nutritional therapy. Further longitudinal studies are indicated to confirm this proposed cytokine–leptin hypothesis in pulmonary cachexia which may then open a novel approach to combat this significant comorbidity in COPD.

References
